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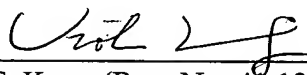
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Sir:

Pursuant to 37 C.F.R. §1.125(a), Applicants submit herewith a substitute specification. Applicants submit that the number or nature of the amendments made to the specification would make it difficult to arrange the paper for printing unless a substitute application is provided. No new matter is added by way of the substitute application. The changes in the specification are purely formatting or correcting to proper English. A marked-up version of the specification showing all the changes made is enclosed herewith separately.

Respectfully submitted,

Date: January 28, 2005



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COMPOSITIONS AND METHODS FOR DIAGNOSIS AND THERAPY OF CANCER

FIELD OF THE INVENTION

The present invention relates to a method for improved diagnosis of cervical
 5 lesions based on detection of gene products encoded by the INK4a gene locus.
 According to the present invention, an improvement in diagnosis may be achieved
 by assessing the presence or absence or the level of overexpression of at least
 two different gene products encoded by the INK4a gene locus. Especially the
 discrimination of p16INK4a overexpressing metaplasias from neoplastic or
 10 preneoplastic p16INK4a overexpressing lesions may be improved by
 determination of the level of other gene-products encoded by the INK4a locus
 such as p14ARF molecules in biological samples in the course of cytological
 testing procedures. The method thus enables for reduction of false positive results
 in the p16INK4a based detection of anogenital lesions in cytological testing
 15 procedures. Furthermore the present invention relates to peptides derived from
 cell cycle regulatory proteins, the expression of which is altered in association with
 tumors in individuals. These peptides according to the present invention may be
 used for detection and therapy of tumors. For detection purposes, the peptides
 may for example be used to detect antibodies directed against said peptides. In
 20 therapeutic respect the peptides may be used for immunotherapy or vaccination
 approaches. In therapeutic and diagnostic respect the peptides may be used in
 combination with one or more peptides derived from tumor associated proteins.

BACKGROUND OF THE INVENTION

The detection of the overexpression of p16INK4a in biological samples has proven
 25 as a useful marker in the detection of anogenital lesions such as carcinoma of the
 uterine cervix (see WO00/01845; Klaes et al., Int. J. Cancer: 92, 276-284 (2001)).
 The method based on p16INK4a-specific immuno-chemical staining allows for a
 sensitive and specific identification of dysplastic cells in tissue section and in
 cytological samples.

30 In immuno-histochemical examinations of tissues, dysplastic and neoplastic cells
 can be stained using a p16INK4a specific antibody mediated staining procedure.

The histological diagnosis of neoplastic lesions can thus be supported by a staining procedure based on a molecular marker characteristic for transformation of cells in anogenital lesions. The diagnosis, whether or not cells are neoplastic, in these procedures is not solely based on the p16INK4a specific staining, but does
5 also rely on the histological information.

This is due to the fact, that in about 20-30% of samples metaplastic cells show some immunoreactivity with p16 INK4a specific antibodies, and thus are stained in the course of the procedures. Yet the staining pattern yielded from these metaplastic cells differs from the pattern, which renders from neoplastic lesions.
10 Metaplastic cells give rise to a patchy or focal staining pattern, whereas neoplastic lesions give rise to diffuse staining pattern. Moreover, the staining intensities of metaplastic cells are predominantly less than that of neoplastic cells.

The common methods used in screening tests for the early detection of dysplasias and/or neoplasias do not employ histology based tests, but do rather rely on
15 cytological testing procedures. Yet especially in cases, when there is no histological information available concerning the architecture of tissues, such as for example in cytological examinations, testing for p16INK4a overexpression alone may lead to false positive results. This is due to the fact that those fractions of metaplastic cells expressing p16INK4a at detectably elevated levels may not be
20 differentiated by means of a histologic criteria.

The percentage of cells showing overexpression of p16INK4a increases in the course of emergence of dysplasias. So in neoplastic or preneoplastic stages, when only a restricted population of neoplastic or preneoplastic cells is present in samples, the immunoreactivity of p16INK4a may be weak. This weak
25 immunoreactivity may be of about the level as the level caused by metaplastic cells. In later stages of dysplasias, the overall immunoreactivity of p16INK4a is stronger and so neoplastic lesions are easily discernible from metaplasias even in a cytological testing format. This might lead to cases, where the presence of metaplastic cells expressing p16INK4a might be confused with the presence of
30 neoplastic cells, and thus produces a false positive result.

Especially in screening tests, where the detection of early stages of neoplasias is desirable, this condition is quite unpleasant. This is especially true, as the

p16INK4a based diagnosis has proven to be a valuable tool in histological examinations and the application in cytological based screening procedures would be able to enhance these established procedures.

To reduce false positive results in cytological testing formats and so to further
5 enhance the fidelity of the p16INK4a mediated diagnosis of anogenital lesions, a method for discriminating the metaplasias from neoplastic and dysplastic lesions would be desirable. The problem in the art especially pertains to early stages of neoplasias, when the percentage of cells showing p16INK4a overexpression is still at a level, that might be confused with levels of normally occurring p16INK4a
10 overexpressing proliferating metaplastic cells. Thus useful means for solving the present problem have to involve parameters that characterize early stages of neoplasias of the anogenital tract. Any characteristics of dysplasias and/or neoplasias emerging during the progress of tumorigenesis, thus proving as diagnostic tools for high grade dysplasias, are limited in early stages of
15 tumorigenesis, and are not suitable for the method according to the present invention.

SUMMARY OF THE INVENTION

A method for the discrimination of metaplasias from neoplastic and preneoplastic lesions is provided within the embodiments claimed according to the present
20 invention.

For supporting the discrimination of metaplasias from neoplastic lesions in testing procedures based on the overexpression of p16INK4a, a marker molecule which is expressed in neoplastic and/or preneoplastic cells and tissues and not expressed in metaplastic cells, would be desirable.

25 A solution to the problem present in the art is provided by the methods claimed according to this invention. In the course of the experiments leading to the present invention, the inventors have found that a combination of detection of the presence or absence and/or the level of p16INK4a in combination with at least one further expression product encoded by the INK4a gene locus, such as the p14ARF
30 protein, may solve the problem present in the art.

Another aspect of the invention relates to use of peptides derived from ink4a locus encoded molecules.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the specimen of cervix uteri, metaplastic area subjected to an immunochemical staining reaction directed against p16INK4a; the figure shows focal positive staining for p16INK4a; for experimental details see Example 1.

- 5 Figure 2 shows the specimen of cervix uteri, metaplastic area subjected to an immunochemical staining reaction directed against p14ARF; the figure shows negative staining for p14ARF; for experimental details see Example 1.

Figure 3 shows the specimen of cervix uteri, dysplastic area (CIN III) subjected to a immunochemical staining reaction directed against p16INK4a; the figure shows
10 diffuse positive staining for p16INK4a; for experimental details see Example 1.

Figure 4 shows the specimen of cervix uteri, dysplastic area (CIN III) subjected to an immunochemical staining reaction directed against p14ARF; the figure shows positive nuclear staining for p14ARF; for experimental details see Example 1.

- Figure 5 shows the cytological specimen of the cervix uteri containing dysplastic
15 area (CIN III) subjected to an sequential immunochemical double staining reaction employing antibodies directed against p14ARF and p16^{INK4a}; the figure shows positive nuclear staining for p14ARF in brown and positive cytoplasmic and nuclear staining for p16^{INK4a} in red; for experimental details see Example 6.

DETAILED DESCRIPTION OF THE INVENTION

- 20 A promising approach for therapy of carcinomas could be based on removal of affected cells or tissues by the immune system of individuals. Analogous to the way the immune system is able to fight infections by micro-organisms etc., certain properties of transformed tumor cells and tissues may be used to elicit an immune response directed against these cells and tissues.
- 25 It is well known in the art that tumors are associated with alterations in molecular properties in cells. Commonly tumors are accompanied by the expression of altered polypeptides or by the expression of non-wild type levels of proteins. This is especially true for cell cycle regulatory proteins.

During the experiments leading to the present invention the inventors have found that peptides derived from cell cycle regulatory proteins, which are expressed in association with tumors, may be used for detection and therapy of tumors. Especially useful are the expression products encoded by the INK4a gene locus.

- 5 The expression products encoded by the INK4a locus are known to show non wild-type expression patterns in association with cancers. It is especially known, that the 16 kDa product p16^{INK4a} encoded by said gene is either overexpressed or not at all expressed in a large number cancers. Furthermore it has been reported, that other expression product encoded by that locus by an alternative reading
10 frame such as p14ARF, which also functions as a cell cycle regulatory protein, albeit as member of a different regulatory pathway, is also overexpressed in association with cancers.

The inventors of the compounds and methods disclosed herein could show that immunogenic peptides may be derived from the expression products encoded by
15 the INK4a locus. These products furthermore may be used to elicit a specific immune response directed against tumor cells expressing the INK4a encoded gene products.

The subject matter specified in the claims of the present patent application is suitable to overcome the disadvantages of therapy and diagnosis of cancers as
20 present in the art. The inventors identified a number of particular peptides, which are suitable for the generation of immune response against the tumor associated proteins and their degradation products. These immunogenic peptides known to show non-wild type expression in cells affected by tumors provide useful tools for the therapy and diagnosis of said diseases.

- 25 The present invention in one aspect relates to peptides and nucleic acids encoding these peptides, which derived from tumor associated proteins, and which may be used to elicit an immune response against said tumors. The peptides are derived from cell cycle regulatory proteins, which show non wild-type expression in association with tumors.

- 30 A second aspect of the present invention is a method for treatment of tumors using the peptides and nucleic acids disclosed herein. The method is based on the generation of an immune response in individuals directed against cells expressing

the respective peptides. The method may be applied as preventive or therapeutic means to fight tumors.

A third aspect of the present invention is a method for diagnosis of tumors based on the detection of antibodies directed against the peptides disclosed herein in
5 individuals. The detection of the antibodies may for example be performed using the disclosed peptides. Otherwise the antibodies may be detected using specific antibodies directed against them.

A fourth aspect of the present invention relates to a pharmaceutical composition for use in the therapeutic method according to the present invention. The
10 pharmaceutical composition is characterized in that it contains at least one of the peptides disclosed herein. Furthermore, the pharmaceutical composition may comprise one or more further peptides derived from a protein, that is associated with cancer.

A fifth aspect of the present invention relates to a kit for the detection of the
15 antibodies directed against the inventive peptides in individuals. The kit may for example contain the respective peptides or antibodies for the detection of the antibodies. The kit may for example be used for diagnostic purposes to detect the presence or absence of a tumor or in the course of immunotherapy to monitor the therapeutic effect. Furthermore a kit according to the present invention may also
20 comprise positive and/or negative control samples.

In a further aspect, the present invention relates to a method for discrimination of neoplastic, preneoplastic and/or dysplastic lesions from metaplasias comprising p16INK4a overexpressing cells, in biological samples in a cytological testing procedure based on the detection of the presence or absence of cells expressing
25 INK4a gene locus encoded gene-products in said biological samples. INK4a gene locus encoded gene-products useful for the method disclosed herein are gene-products such as e.g. p14ARF. In one embodiment of the invention, p14ARF protein or mRNA may serve as a marker for discrimination of metaplasias from early neoplastic or preneoplastic lesions in samples.

30 The present invention provides peptides and nucleic acids coding therefore, which are expressed in association with tumors, for the use in therapy and diagnosis of

disorders such as cancer. The peptides are derived from cell cycle regulatory proteins, which show an altered expression in association with cancer.

The INK4a gene locus as used in the context of the present invention is located on chromosome 9p21. The locus is alternatively nominated e.g. the INK4a/ARF locus,
 5 CDKN2A gene, the MTS1 locus. The gene locus dealt with herein is that gene locus, that in one aspect encodes the gene product p16^{INK4a} protein, but is also known to encode for other gene product in alternative reading frames.

Altered expression as used in the context of the present invention may comprise for example any non wild-type expression of a nucleic acid. The expression may
 10 differ from the wild type expression either in the level of the expression (elevated expression level or reduced expression level) or the expression product (mutated expression product). A mutated expression product differs from the wild type expression product in sequence or in any structural parameter such as modification etc. The wild-type expression shall be understood to be the
 15 expression found in a normal control tissue.

An expression product according to the present invention may be any molecule transcribed from a gene or any molecule translated from such a transcript. Thus expression product as used in the context of the present invention may comprise polynucleotides such as e.g. DNA or RNA and polypeptides such as proteins,
 20 proteoglycans, peptides etc. Expression product as used in the context of the present invention shall comprise any transcript of a gene locus in forward or reverse direction including any reading frames, splicing variants or alternative promoters. Expression products as used herein shall thus comprise any alternative products encoded by the nucleic acids of a particular gene locus.

25 In one embodiment of the invention, the cell cycle regulatory proteins are encoded by the INK4a gene. The cell cycle regulatory proteins as disclosed herein comprise any proteins encoded by alternative reading frames, arising from alternative splicing or usage of alternative promoter sequences of said gene. In one embodiment of the present invention the proteins exhibit molecular weights of
 30 about 5 to 40 kDa or any value in between and preferably of about 10 to 20 kDa or any value in between and most preferably of about 14 to about 19 kDa or any value in between respectively. The cell cycle regulatory proteins for use in the

context of the present invention are different from p16INK4a cyclin dependent kinase inhibitor in at least one property selected from the group consisting of molecular weight, amino acid sequence, biological function and immunoreactivity.

In one embodiment of the present invention, the immunogenic peptides that may be derived from a protein may be predicted as described by Rammensee et al, "MHC ligands and peptide motifs: 1st listing", Immunogenetics **41**, 178-228, 1995 and Rammensee, et al., "MHC ligands and peptide motifs" Bioscience 1997, which methods shall be included herein by reference. The methods are exemplified below in the examples.

Peptides derived from the cell cycle regulatory proteins as used in the context of the present invention may comprise the respective full length proteins, fragments thereof, or peptides of any length. In certain embodiments of the present invention the peptides derived from cell cycle regulatory proteins showing altered expression in association with cancers may be e.g. at least one or more peptides selected from the peptides given below.

In one embodiment of the present invention, the peptides derived from the INK4a expression products are 15-mer peptides. The 15-mer peptides are for example selected from the group consisting of the peptides :

G H D D G Q R P S G G A A A A (SEQ ID NO: 1)

V V H I P R L T G E W A A P G (SEQ ID NO: 2)

V H I P R L T G E W A A P G A (SEQ ID NO: 3)

R R P G H D D G Q R P S G G A (SEQ ID NO: 4)

In this embodiment, the peptides are preferably presented by MHC class II molecules.

In another embodiment, the peptides may be HLA-A1 restricted nonamer polypeptides selected from a group consisting of the following:

H D D G Q R P S G (SEQ ID NO: 5)

A A E L G P G G G (SEQ ID NO: 6)

R R C P G G L P G (SEQ ID NO: 7)

G H D D G Q R P S (SEQ ID NO: 8)

G G E N M V R R F (SEQ ID NO: 9)

In yet another embodiment, the peptides are HLA-A3 restricted nonamers selected from a group consisting of:

- 5 Q L R R P R H S H (SEQ ID NO: 10)
 M V R R F L V T L (SEQ ID NO: 11)
 V L M L L R S Q R (SEQ ID NO: 12)
 R L G Q Q P L P R (SEQ ID NO: 13)
 R V R V F V V H I (SEQ ID NO: 14)
 10 T L R I R R A C G (SEQ ID NO: 15)
 R L T G E W A A P (SEQ ID NO: 16)

In one further embodiment, the peptides may be HLA A*0201 restricted nonamers selected from a group comprising the following peptides:

- M V R R F L V T L (SEQ ID NO: 17)
 15 A A V A L V L M L (SEQ ID NO: 18)
 A V A L V L M L L (SEQ ID NO: 19)
 L M L L R S Q R L (SEQ ID NO: 20)
 H I M G R G R C V (SEQ ID NO: 21)
 A A P G A P A A V (SEQ ID NO: 22)
 20 G A P A A V A L V (SEQ ID NO: 23)

(Poly)peptides as used according to the present invention may comprise amino acid chains of any length, including full length proteins, wherein the amino acid residues are linked by covalent peptide bonds.

Tumor associated peptides as disclosed in the context of the present invention
 25 shall comprise polypeptides of lengths of at least 4 amino acids. The peptides may for example comprise 4 to 50 amino acids or any number of amino acids in

between. In another embodiment of the present invention, the peptides may comprise polypeptides with more than 50 amino acids. These polypeptides for use in the context of the present invention may for example comprise 50, 100, 500, 750, 1000 amino acids or any number of amino acids in between and may
5 comprise proteins, or fragments thereof, and/or fusion- or chimeric proteins comprising on or more additional heterologous sequences. The additional sequences may be derived from the native protein or may be heterologous, and such sequences may (but need not) be immunoreactive and/or antigenic. As detailed below, such polypeptides may be isolated from tumor tissue or prepared
10 by synthetic or recombinant means.

As used herein, a polypeptide exhibiting biological properties of the tumor associated peptides is understood to be a polypeptide having at least substantially the same immunogenic properties, i.e. is still capable of binding an antibody directed against said polypeptide, e.g. comprises an immunogenic portion of said
15 polypeptide.

Peptides disclosed herein are immunogenic polypeptides. This requires that the polypeptides may stimulate immune responses in host organisms either in the form the polypeptides adopt in their natural environment and/or especially in the form the polypeptides adopt after processing by the cellular antigen processing
20 and presenting machinery.

Immunogenic portion as used above is a portion of a protein, which is recognized by a B-cell and/or T-cell surface antigen receptor. The immunogenic portions comprise at least 4 amino acid residues, at least 10 amino acid residues or at least 15 amino acid residues of the protein disclosed herein. In one embodiment of the
25 present invention, particular domains of the protein, such as for example transmembrane domains or N-terminal leader sequences have been deleted.

The immunogenic portions according to the present invention react with antisera or specific antibodies in the same or nearly same intensity as the native full length proteins. The immunogenic portions are generally identified using the techniques
30 well known in the art. Possible techniques are for example screening of the polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones.

In certain embodiments of the present invention, polypeptides may comprise fusion or chimeric polypeptides containing sequences disclosed herein. Fusion proteins comprise the polypeptide according to the present invention together with any second and further polypeptides, such as e.g. one or more polypeptides of the same sequence or of another sequence. Heterologous polypeptides may comprise e.g. enzymes, receptor molecules, antigens, antigenic or immunogenic epitopes or fragments, antibodies or fragments thereof, signalling polypeptides or signal transducing polypeptides, labelled polypeptides etc.. The immunogenic protein may for example be capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (see, for example, Stoute et al. *New Engl. J. Med.*, 336:86-91 (1997)).

In one embodiment of the invention, the fusion peptides may be constructed for enhanced detection or purification of the polypeptides, or of complexes of the polypeptides with the respective immunological entities according to the present invention. For the purpose of purification tags, such as e.g. his-tags, myc-tags etc. may be added to the polypeptides. For the purpose of detection antigenic portions, enzymes, chromogenic sequences etc. may be fused to the polypeptides. The fusion proteins of the present invention may (but need not) include a linker peptide between the first and second polypeptides.

A peptide linker sequence may be employed to separate the first and the second polypeptides by a distance sufficient to ensure, that each polypeptide folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion protein using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes.

Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea et al., *Gene* 40:39-46, 1985; Murphy et al., *Proc. Natl. Acad.*

Sci. USA 83:8258-8262, 1986; U.S. Pat. No. 4,935,233 and U.S. Pat. No. 4,751,180. The linker sequence may be from 1 to about 50 amino acids in length. Peptide sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the
5 functional domains and prevent steric interference.

The term tumor associated (poly)peptides according to the present invention may also comprise variants of the native proteins. These variants may differ from the native protein in one or more alterations such as substitutions, deletions, additions and/or insertions. The immunoreactivity of the variants according to the present
10 invention is not substantially diminished compared to the native proteins. In a preferred embodiment of the invention the immunoreactivity and or activity is diminished less than 50% and in a more preferred embodiment the immunoreactivity and or activity is diminished less than 20 % compared to the native polypeptides.

15 The variants according to the present invention comprise e.g. conservative substitutions, so that the amino acids changed are substituted for amino acids with similar properties. The properties concerned may include polarity, charge, solubility, hydrophobicity, hydrophilicity and/or amphipathic nature of the amino acid residues.

20 The variants according to the invention may also comprise additional terminal leader sequences, linkers or sequences, which enable synthesis, purification or stability of the polypeptides in an easier or more comfortable way.

Variants of the polypeptides according to the present invention may be produced by means of conventional molecular biological processes (see, e.g., Sambrook et
25 al., 1989, Molecular Cloning, A Laboratory Manual, 2nd edition Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY)

The tumor associated polypeptides disclosed herein may be generated by synthetic means. In particular, synthetic polypeptides having fewer than about 100 amino acids, and generally fewer than about 50 amino acids, may be generated
30 using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where

amino acids are sequentially added to a growing amino acid chain (see, for example, Merrifield, J. Am. Chem. Soc. 85:2149-2146, 1963). Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, Calif.), and may be
5 operated according to the manufacturer's instructions.

According to the present invention, polypeptides may be represented by nucleic acids coding for said polypeptides. These nucleic acids may for example be used for the *in situ* expression of the respective polypeptides. For the purpose of expression of nucleic acids, the particular nucleic acids may be joined with suitable
10 other nucleic acid sequences, that enable for the cloning and expression of said nucleic acids encoding the tumor associated polypeptides.

Nucleic acid molecules according to the present invention may comprise polynucleotides or fragments thereof. Preferred polynucleotides may comprise at least 20 consecutive nucleotides, preferably at least 30 consecutive nucleotides and
15 more preferably at least 45 consecutive nucleotides of the sequences. The nucleic acids according to the present invention may also be complementary or reverse complementary to any of said polynucleotides. Polynucleotides may for example include single-stranded (sense or antisense) or double-stranded molecules, and may be DNA (genomic, cDNA or synthetic) or RNA. RNA molecules comprise as
20 well HnRNA (containing introns) as mRNA (not containing introns). According to the present invention the polynucleotides may also be linked to any other molecules, such as support materials or detection marker molecules, and may, but need not, contain additional coding or non-coding sequences.

A nucleic acid sequence encoding one or more peptides of the present invention is
25 constructed using known recombinant DNA techniques to assemble separate nucleic acid sequences encoding the first and second polypeptides into an appropriate expression vector. The 3' end of a nucleic acid sequence encoding the first polypeptide is ligated, with or without a peptide linker, to the 5' end of a nucleic acid sequence encoding the second polypeptide ensuring the appropriate
30 reading frames of the sequences to permit mRNA translation of the two nucleic acid sequences into a single fusion protein that retains the biological activity (antigenicity) of both the first and the second polypeptides.

The nucleotide sequences according to the present invention may be joined to a variety of other nucleic acid sequences using the known recombinant DNA techniques. The sequences may for example be cloned into any of a variety of cloning vectors, such as plasmids, phagemids, lambda phage derivatives and
5 cosmids. Furthermore, vectors such as expression vectors, replication vectors, probe generation vectors and sequencing vectors may be joined with the sequences disclosed herein. Sequences of special interest, which could be cloned to the nucleic acids according to the present invention, are for example non coding sequences and regulatory sequences including promoters, enhancers and
10 terminators.

In a preferred embodiment, polynucleotides may be formulated such that they are able to enter mammalian cells and to be expressed in said cells. Such formulations are especially useful for therapeutic purposes. The expression of nucleic acid sequences in target cells may be achieved by any method known to
15 those skilled in the art. The nucleic acids may for example be joined to elements that are apt to enable their expression in a host cell. Such elements may comprise promoters or enhancers, such as CMV-, SV40-, RSV-, metallothionein I- or polyhedrin-promoters respectively CMV- or SV40-enhancers. Possible methods for the expression are for example incorporation of the polynucleotides into a viral
20 vector including adenovirus, adeno-associated virus, retrovirus, vaccinia virus or pox virus. Viral vectors for the purpose of expression of nucleic acids in mammalian host cells may comprise pcDNA3, pMSX, pKCR, pEFBOS, cDM8, pCEV4 etc.. These techniques are known to those skilled in the art.

Other formulations for administration in therapeutic purposes include colloidal
25 dispersion systems such as for example macromolecule complexes, microspheres, beads, micelles and liposomes.

For the manipulation in prokaryotic cells by means of genetic engineering the nucleic acid molecules of the invention or parts of these molecules can be introduced into plasmids allowing a mutagenesis or a modification of a sequence
30 by recombination of DNA sequences. By means of conventional methods (cf. Sambrook et al., supra) bases may be exchanged and natural or synthetic sequences may be added. In order to link the DNA fragments with each other

adapters or linkers may be added to the fragments. Furthermore, manipulations may be performed that provide suitable cleavage sites or that remove superfluous DNA or cleavage sites. If insertions, deletions or substitutions are possible, in vitro mutagenesis, primer repair, restriction or ligation may be performed. As analysis
5 method usually sequence analysis, restriction analysis and other biochemical or molecular biological methods may be used.

A nucleic acid sequence encoding a fusion protein of the present invention is constructed using known recombinant DNA techniques to assemble separate nucleic acid sequences encoding the first and second polypeptides into an
10 appropriate expression vector. The 3' end of a nucleic acid sequence encoding the first polypeptide is ligated, with or without a peptide linker, to the 5' end of a nucleic acid sequence encoding the second polypeptide ensuring the appropriate reading frames of the sequences to permit mRNA translation of the two nucleic acid sequences into a single fusion protein that retains the biological activity of
15 both the first and the second polypeptides.

The tumor associated polypeptides of the present invention, and nucleic acids encoding such polypeptides, may be isolated from tumor tissue using any of a variety of methods well known in the art. Nucleic acid sequences corresponding to a gene (or a portion thereof) encoding one of the inventive tumor polypeptides
20 may be isolated from a tumor cDNA library using a subtraction technique. Partial nucleic acid sequences thus obtained may be used to design oligonucleotide primers for the amplification of full-length nucleic acid sequences from a human genomic nucleic acid library or from a tumor cDNA library in a polymerase chain reaction (PCR), using techniques well known in the art (see, for example, Mullis et
25 al., Cold Spring Harbor Symp. Quant. Biol. 51:263, 1987; Erlich ed., PCR Technology, Stockton Press, NY, 1989). For this approach, sequence-specific primers may be designed based on the nucleotide sequences provided herein and may be purchased or synthesized.

The ligated nucleic acid sequences encoding the inventive polypeptides are
30 operably linked to suitable transcriptional or translational regulatory elements known to the person skilled in the art. The regulatory elements responsible for expression of nucleic acid may be located e.g. 5' to the nucleic acid sequence

encoding the first polypeptides, within the coding sequences or 3' to the nucleic acid sequences encoding the first or any further polypeptide. Stop codons required to end translation and transcription termination signals are present 3' to the nucleic acid sequence encoding the second polypeptide.

- 5 The (poly)peptides as claimed according to the present invention are isolated. This means that the molecules are removed from their original environment. Naturally occurring proteins are isolated if they are separated from some or all of the materials, which coexist in the natural environment. Polynucleotides are isolated for example if they are cloned into vectors.
- 10 In certain preferred embodiments, described in more detail below, the polypeptides disclosed herein are prepared in an isolated, substantially pure form (i.e., the polypeptides are homogenous as determined by amino acid composition and primary sequence analysis). Preferably, the polypeptides are at least about 90% pure, more preferably at least about 95% pure and most preferably at least 15 about 99% pure. The substantially pure polypeptides may for example be employed in pharmaceutical compositions.

In certain embodiments of the present invention, especially for the detection of specific antibodies directed against tumor associated polypeptides, these polypeptides themselves may be employed. Immunogenic portions as used herein 20 is a portion of a protein, that is recognized by a B-cell and/or T-cell surface antigen receptor. The immunogenic portions comprise at least 5 amino acid residues, more preferably at least 10 amino acid residues and most preferably at least 15 amino acid residues of the tumor associated polypeptides according to the present invention.

- 25 Immunogenic portions useful for the detection of specific antibodies may be provided as oligopeptides or as part of larger proteins. This is dependent on the embodiment of the invention. Where antibodies are to be detected, the antigenic epitopes may either be primary structures of polypeptides or the epitopes may be built by complex arrangements of tertiary structures of polypeptides. Concerning 30 cells directed against specific tumor associated peptides the relevant immunogenic portions are merely short fragments of peptides with a length of

about 10 – 20 amino acids. Thus depending on the particular detection method the immunogenic portions have to be chosen.

Another aspect of the present invention relates to binding agents, antibodies and immunological entities specifically recognizing the peptides derived from proteins associated with tumors. In this respect, the present invention provides binding agents such as antibodies and antigen-binding fragments that specifically bind to the peptides derived from INK4a encoded proteins showing altered expression patterns in tumors.

The term binding agent comprises a variety of substances such as oligopeptides, antibodies, peptidomimetic molecules comprising antigen binding oligopeptides, nucleic acids, carbohydrates, organic compounds, etc. Antibody according to the present invention relates to antibodies which consist of polyclonal antibodies, essentially of pooled monoclonal antibodies with different epitopic specificities, as well as distinct monoclonal antibody preparations. Monoclonal antibodies are made using an antigen containing fragments of the polypeptides of the invention by methods well known to those skilled in the art (see, e.g., Köhler et al., Nature 256 (1975), 495). As used herein, the term “antibody” (Ab) or “monoclonal antibody” (Mab) is meant to include intact molecules as well as antibody fragments (such as, for example, Fab and F(ab')₂ fragments) which are capable of specifically binding to protein. Fab and f(ab')₂ fragments lack the Fc fragment of intact antibody, clear more rapidly from the circulation, and may have less non-specific tissue binding than an intact antibody. (Wahl et al., J. Nucl. Med. 24: 316-325 (1983)). Thus, these fragments are preferred, as well as the products of a FAB or other immunoglobulin expression library. Moreover, antibodies of the present invention include chimerical, single chain, and humanized antibodies.

For certain purposes, e.g. diagnostic methods, the antibody or binding agent of the present invention may be detectably labelled, for example, with a radioisotope, a bioluminescent compound, a chemiluminescent compound, a fluorescent compound, a metal chelate, or an enzyme. Furthermore any method suitable for the detection of the intermolecular interaction may be employed.

The antibody or antigen-binding agent is said to react specifically, if it reacts at a detectable level with a peptide disclosed herein, and does not significantly react

with other peptides. The antibodies according to the present invention may be monoclonal or polyclonal antibodies. Other molecules capable of binding specifically may be for example antigen-binding fragments of antibodies such as Fab fragments, RNA molecules or polypeptides. According to the present
5 invention binding agents may be used isolated or in combination. By means of combination it is possible to achieve a higher degree of sensitivity.

The antibodies useful for the methods according to the present invention may comprise further binding sites for either therapeutic agents or other polypeptides or may be coupled to said therapeutic agents or polypeptides. Therapeutic agents
10 may comprise drugs, toxins, radio-nuclides and derivatives thereof. The agents may be coupled to the binding agents either directly or indirectly for example by a linker or carrier group. The linker group may for example function in order to enable the coupling reaction between binding agent and therapeutic or other agent or the linker may act as a spacer between the distinct parts of the fusion molecule.
15 The linker may also be cleavable under certain circumstances, so as to release the bound agent under said conditions. The therapeutic agents may be covalently coupled to carrier groups directly or via a linker group. The agent may also be non-covalently coupled to the carrier. Carriers that can be used according to the present invention are for example albumins, polypeptides, polysaccharides or
20 liposomes.

The antibody used according to the present invention may be coupled to one or more agents. The multiple agents coupled to one antibody may be all of the same species or may be several different agents bound to one antibody.

The invention also relates to a transgenic non-human animal such as transgenic
25 mouse, rats, hamsters, dogs, monkeys, rabbits, pigs, C. elegans and fish such as torpedo fish comprising a nucleic acid molecule or vector of the invention, preferably wherein said nucleic acid molecule or vector may be stably integrated into the genome of said non-human animal, preferably such that the presence of said nucleic acid molecule or vector leads to the expression of the inventive cell
30 cycle regulatory protein derived polypeptide (or related polypeptide) of the invention, or may otherwise be transiently expressed within the non-human animal. Said animal may have one or several copies of the same or different

nucleic acid molecules encoding one or several forms of the inventive polypeptide. This animal has numerous utilities, including as a research model for the development of therapies, treatment, etc. for tumors. Accordingly, in this instance, the non-human mammal is preferably a laboratory animal such as a mouse or rat.

- 5 It is one aspect of the present invention to detect tumors on the basis of the detection of immunological entities directed against the peptides disclosed herein.

The detection as used herein may also comprise the detection of immunological entities disclosed herein together with immunological entities directed against other tumor associated peptides. These peptides may e.g. comprise p13,5, p14,
10 p15, p16, p19, p21, p27, p53, Rb protein, osteopontine, grp, renal dipeptidase, id1, mcm5, mcm2, cdc6, MDM2. The peptides furthermore may for example be derived from peptides encoded by the genes disclosed in the documents WO9904265A2, WO0149716A2, WO0055633A2 and WO0142792A2, which shall be included by reference herein. In one embodiment of the invention a set of polypeptides may be
15 used for the detection of antibodies. The set may be a combination of relevant peptides in solution, in cases, where information about the presence of immunological entities in general is sought. In contrast, when information about the presence or absence of particular peptides within a set of peptides is sought, the peptides may for example be tested each as a single, simultaneously in
20 multiple testing reactions. Such experiments may for example be carried out in form of multi-well tests or using peptide arrays etc.

The method for detection of disorders according to the present invention thus may employ the detection of immunological entities directed against one single peptide or the detection of a set of immunological entities. The use of a multiplicity of
25 potential peptides raises the probability to detect the presence of a particular disorder, and may furthermore give additional information useful in stratification of a disorder, in monitoring the disease course or in assessment of prognosis concerning the disease course.

Immunological entities as used in the context of the present invention shall comprise
30 any components of the mammalian immune system, that are able to specifically react with an antigenic epitope. Such immunological entities may comprise for

example antibodies, all immunoglobulins, such as e.g. IgG, IgM, IgA, IgE, IgD, specific CD8+ T-cells or specific T-helper cells.

Diagnosis as used in the context of the present invention may comprise determining the presence or absence and/or the level of specific immunological entities directed
5 against particular tumor associated peptides in a sample, and assessing diagnosis from said presence or absence and/or level of immunological entities specifically directed against said tumor associated polypeptides.

Based upon the determined presence or absence and/or the levels of immunological entities specifically directed against particular tumor associated
10 peptides in the samples individuals can be subdivided into subgroups. The subgroups may be created according to scientific or clinical data, such as e.g. survival, recurrence of disease, frequency of metastases etc., related to the presence or absence and/or levels of particular tumor associated peptides in samples of tissues affected with a particular disorder, of tissues being in question
15 of being affected with a particular disorder or of tissues at risk of being affected with a particular disorder.

Based upon these subgroups, an assessment of prognosis may be done. According to the subgroups the therapy of the individuals affected by the disorders (e.g. tumors) may be tailored.

20 Monitoring may comprise detecting the presence or absence and/or the level of immunological entities specifically directed against tumor associated (poly)peptides in samples taken at different points in time and determining the changes in said levels or presences or absences. According to said changes the course of the disease can be followed. E.g. the occurrence of immunological
25 entities directed against tumor associated (poly)peptides, that have not been present at an earlier time-point may be indicative of the progression of evolution of the affected tissue. The course of the disease may be used to select therapy strategies for the particular individual.

Monitoring may also pertain to the monitoring of the therapeutic effect in the
30 course of an immunotherapy according to the present invention'.

Another aspect of diagnosis and monitoring of the disease course according to the present invention may comprise the detection of minimal residual disease. This

may comprise for example the detection of presence and/or level of immunological entities specifically directed against said tumor associated (poly)peptides, that have not been present in earlier examinations in one or more body samples following initial therapy of an individual once or at several time points. According to
5 the presence and/or level of immunological entities specifically directed against tumor associated polypeptides detected in the samples, one may select a suitable therapy for the particular individual.

Furthermore, the diagnostic method may be carried out to detect disseminated tumor cells in biological samples as MRD diagnosis of minimal residual disease. For
10 this purpose the detection of the level of immunological entities or the presence of immunological entities, specific for particular peptides, that have not been detected in prior examinations, may be performed.

A sample according to the method of the present invention may comprise any sample comprising immunological entities as defined above. Samples may
15 comprise samples of clinical relevance, such as e.g. blood, serum, plasma, secretions, smears, body fluids, urine, semen, stool, sputum, bile, biopsies, cell- and tissue-samples. Biopsies as used in the context of the present invention may comprise e.g. resection samples of tumors, tissue samples prepared by endoscopic means or needle biopsies of organs.

20 Such samples may comprise for example intact cells, lysed cells or any liquids containing antibodies, immunoglobulins or cells specifically directed against tumor associated peptides. Even solids, to which cells, cell fragments or antigen binding polypeptides, such as antibodies or immunoglobulins may adhere, or may be fixed, may be samples according to the method disclosed herein. The method for
25 detection of the level of the immunological entities according to the present invention is any method, which is suited to detect very small amounts of specific immunological entities in biological samples. The detection reaction according to the present invention is a detection either on the level of antibodies or on the level of cells specific for particular antigens.

30 The compositions and methods according to the present invention may be applied to any eukaryotic organism exhibiting an immunologic defence system. The eukaryotic organisms are for example animals of agricultural value such as pigs,

cows, sheep, etc., companion animals, such as cats, dogs, horses etc., animals employed in research purposes such as mice, rats, rabbits, hamsters etc. or humans.

The methods disclosed herein may be applied generally to proliferative disorders.

5 Proliferative disorders according to the present invention comprise diseases characterized by abnormal growth properties of cells or tissues compared to the growth properties of normal control cells or tissues. The growth of the cells or tissues may be for example abnormally accelerated, decelerated or may be regulated abnormally. Abnormal regulation as used above may comprise any form

10 of presence or absence of non wild-type responses of the cells or tissues to naturally occurring growth regulating influences. The abnormalities in growth of the cells or tissues may be for example neoplastic or hyperplastic.

In one embodiment, the cell proliferative disorders are tumors. Tumors may comprise tumors of the head and the neck, tumors of the respiratory tract, tumors

15 of the gastrointestinal tract, tumors of the urinary system, tumors of the reproductive system, tumors of the endocrine system, tumors of the central and peripheral nervous system, tumors of the skin and its appendages, tumors of the soft tissues and bones, tumors of the lymphopoietic and hematopoietic system etc.. Tumors may comprise for example neoplasms such as benign and malignant

20 tumors, carcinomas, sarcomas, leukemias, lymphomas or dysplasias. In a preferred embodiment the tumor is for example cancer of the of the head and the neck, cancer of the respiratory tract, cancer of the gastrointestinal tract, cancer of the skin and its appendages, cancer of the central and peripheral nervous system, cancer of the urinary system, cancer of the reproductive system, cancer of the

25 endocrine system, cancer of the soft tissues and bone, cancer of the hematopoietic and lymphopoietic system.

In certain embodiments of the invention, the carcinoma is cervical cancer, colon cancer, gastric cancer, breast cancer, bladder cancer, lung cancer etc.

The detection may be carried out in solution or using reagents fixed to a solid

30 phase. Solid phases may comprise beads of a variety of materials, such as e.g. agarose, dextrane polymers, polystyrene, silica, etc. or surfaces of suitable materials such as e.g. polystyrene, glass, agarose, protein, dextran etc. coated

surfaces etc.. The detection of one or more immunological entities, such as immunoglobulins or cells carrying specific antigen recognizing epitopes, with different antigen binding specificities may be performed in a single reaction mixture or in two or more separate reaction mixtures. Alternatively the detection
5 reactions for several immunological entities may for example be performed simultaneously in multi-well reaction vessels.

The immunological entities specifically recognizing particular tumor associated peptides may be detected using reagents that specifically recognise these immunological entities alone or in complex with their respective antigen (e.g.
10 antibodies), or reagents, that are specifically recognized by the immunological entities themselves (e.g. the antigen). In one embodiment, the antigen may be fused to another polypeptide, so as to allow binding of the antigen by the immunological entity in question and simultaneously binding of the second part of the fusion protein by another (labelled) antibody for the detection. The detection
15 reaction for the immunological entities may comprise one or more reactions with detecting agents either recognizing the initial entities or recognizing the prior molecules used to recognize the immunological entities.

The detection reaction further may comprise a reporter reaction indicating the presence or absence and/or the level of the immunological entities. The reporter
20 reaction may be for example a reaction producing a coloured compound, a bioluminescence reaction, a chemiluminescent reaction, a fluorescence reaction, generally a radiation emitting reaction etc.. The detection reactions may for example employ antibodies or binding reagents that are detectably labelled.

Furthermore, the binding of detection molecules to the entities in question may be
25 detected by any measurable changes in physical or physical-chemical properties, such as changes in spectroscopic properties, in magnetic resonance properties etc.. Different immunological entities or immunological entities of different specificities may be recognized by different methods or agents. This may be due to difficulties in detection of several entities, or entities with particular specificities
30 by a particular method. An advantage of the use of different detection techniques for different immunological entities or for immunological entities with different

specificities may for example be, that the different reporter signals referring to different immunological entities could be distinguished.

Generally in a method according to the present invention, the detection of different immunological entities such as the detection of immunoglobulins and the detection
5 of immunocompetent cells may be performed simultaneously. The detection may be e.g. performed simultaneously in one vessel or in different reaction containers or samples. Furthermore the reaction according to the present invention may be performed sequentially one subsequent to the other.

For all detection purposes, optionally the original sample may be concentrated by
10 any suitable means known to those of ordinary skill in the art. Furthermore steps may be involved to selectively extract immunological entities from the sample mixture such as affinity based purification techniques either employing specific antibodies or the respective antigen recognized by the entities in question.

Applicable formats for the detection reaction according to the present invention
15 may be, (reverse) blotting techniques, such as Western-Blot. The blotting techniques are known to those of ordinary skill in the art and may be performed for example as electro-blots, semidry-blots, vacuum-blots or dot-blots. Furthermore immunological methods for detection of molecules may be applied, such as for example immunoprecipitation or immunological assays, such as ELISA, RIA,
20 Elispot assay, lateral flow assays, immuno-cytochemical methods etc.

In one preferred embodiment of the invention, the detection of the level of immunological entities specific for tumor associated peptides is carried out on the level of antibodies. This may be e.g. performed using the specific interaction between the respective tumor associated peptides with the antibodies. The
25 determination of the presence or absence and/or the level of the antibodies may for example be carried out with recombinantly produced tumor associated peptides. The peptides can be used in many different detection techniques for example in western-blot, ELISA or immuno-precipitation. In one embodiment the detection of antibodies is carried out as antibody capture assay (Antibodies A
30 laboratory Manual, Harlow, Ed. et al., Cold Spring Harbor Laboratory 1988).

In another embodiment of the invention, the detection of the specific antibodies is carried out using monoclonal or polyclonal antibodies specifically recognizing the

antigen binding epitope of the first antibodies. For this purpose the above mentioned immunological detection procedures may be applied. In a further embodiment, chimeric antigens may be employed in the detection reaction. Such chimeric antigens may for example comprise fusion proteins combining the antigenic epitope
 5 of a tumor associated polypeptide, recognized by the antibody in question, fused to another antigen, that may be recognized by a detection antibody. The particular antigens within the chimeric polypeptide may be separated by a linker or spacer region.

Any other method for determining the amount of particular antibodies or
 10 immunoglobulins in biological samples can be used according to the present invention.

Generally, the detection of the antibodies according to the present invention may be performed as well in vitro as directly in situ, for example, in the course of an immuno-histochemical or immuno-cytochemical staining reaction.

15 Cells exhibiting specificity for a particular antigen may be detected by any methods suitable for that purpose known to those of ordinary skill in the art. Methods may for example comprise proliferation-assays, cytokine-ELISAs, ELISpot assays, intracellular FACS-staining, PCR-mediated identification of peptide-specific cytokine (or similar) -expressing cells, tetramer-staining, cytotoxicity assays and
 20 DTH-(delayed type hypersensitivity) reactions.

In case of proliferation-assays, induction of peptide-specific T-cell proliferation may be measured by methods known to those of skill in the art. This can be achieved by simple counting of cells, by measuring incorporation of labelled nucleotides into cellular DNA or by measuring level and/or activity of cellular protein(s). Cytokine-
 25 ELISA may comprise identification of peptide-specific cytokine-secreting cells by measuring cytokine levels in supernatant. In the course of an ELISpot assay the number of peptide-specific cytokine (i.e. IFN-g)- secreting cells in a sample is determined. Similarly the intracellular FACS-staining identifies cytokine-expressing cells on the protein level. In contrast (real-time) PCR may be used for identification
 30 of peptide-specific cytokine (or similar) -expressing cells on the transcript level. In the course of a tetramer-staining assay the label is a tetramer-molecule of recombinant MHC-class I molecules, loaded with specific peptide and coupled to a

dye. The tetramer binds to the T-cell receptor. Cytotoxicity assays are a method for identification of cells, that can recognize and kill target cells in a peptide-specific manner. DTH-(delayed type hypersensitivity) reaction is based on the measuring of skin-reaction of vaccinated persons after intradermal (or similar)

5 application of peptide(s).

The method for the detection of immunological entities as disclosed herein may be performed for the purpose of monitoring in the course of immunotherapeutic treatments of individuals. In this respect the presence or absence and or the level of antibodies directed against an inventive peptide in an individual is determined.

10 The determination of the level may be performed using the methods as set forth above. The detection may be performed at several consecutive points in time as to monitor the timely alteration of the level of the immunological entities. The determination may for example be performed daily, weekly, monthly, once a year, or in once in a decade or at any interval in between.

15 Another aspect of the present invention is a testing kit for performing the method according to the present invention. The kit may be for example a diagnostic kit or a research kit.

A kit according to the present invention comprises at least an agent suitable for detecting the immunological entities according to the method disclosed herein.

20 Furthermore, a kit according to present invention may comprise:

a) reagents for the detection of the antibodies or cells specifically recognizing antigens,

b) reagents and buffers commonly used for carrying out the detection reactions as described herein, such as buffers, detection-markers, carrier substances and

25 others, and

c) a sample for carrying out a positive control reaction, that may comprise an antigen or a set of antigens, to which all members of a target population of individuals have antibodies.

The reagent for the detection of the antibodies or cells specifically recognizing
30 antigens may include any agent capable of binding to the antibodies or cells specifically recognizing antigens. Such reagents may include proteins,

polypeptides, nucleic acids, peptide nucleic acids, glycoproteins, proteoglycans, polysaccharides or lipids.

The sample for carrying out a positive control may comprise for example an antigenic peptide or a set of antigenic peptides. Suitable antigens may include
5 antigens, against which a wide percentage of the population has antibodies. Examples of such antigens may for example comprise antigens present in lysed E. coli cells, tetanus antigen, the viral capsid antigen of the Epstein-Barr virus, antigens derived from matrix proteins of Haemophilus influenzae. The antigens may for example be used as a mixture to ensure, that a particular individual
10 actually displays a positive reaction.

In a preferred embodiment of the invention, the detection of the immunological entities directed against particular tumor associated polypeptides is carried out on the level of antibodies. In this embodiment, the binding agent may be for example a tumor associated polypeptide or a fragment thereof, recognized by the
15 respective antibody, or a fusion polypeptide comprising said polypeptide or a fragment thereof. Furthermore, the binding agent may comprise an antibody or a fragment thereof specific for the antibody in question, for the complex of the antibody with the respective tumor associated polypeptide or for an antigenic epitope fused to the polypeptide.

20 In an other embodiment of the test kit, the detection of the immunological entities is carried out on the level of cells specifically recognizing tumor associated polypeptides. In this embodiment of the invention, the reagent for the detection may be for example a tumor associated polypeptide or a fragment thereof, recognized by the respective antibody or T-cell receptor, or a fusion polypeptide
25 comprising said tumor associated polypeptide or a fragment thereof. Furthermore the binding agent may comprise an antibody or a fragment thereof specific for the antibody in question, for the complex of the antibody with the respective polypeptide or for an antigenic epitope fused to the polypeptide.

The tumor associated peptides disclosed herein may be used in vaccines, that
30 may comprise one or more tumor associated polypeptides as disclosed herein and may comprise optionally one or more further tumor associated peptides.

According to the present invention, tumor associated polypeptides that comprise an immunogenic portion may be used for immuno-therapy for the treatment of cancer. Immunotherapy may be broadly classified into either active or passive immunotherapy. In active immunotherapy, treatment relies on the in vivo
5 stimulation of the endogenous host immune system to react against tumours with the administration of immune response-modifying agents (for example, tumour vaccines, bacterial adjuvants, and/or cytokines). A patient may be afflicted with disease, or may be free of detectable disease. Accordingly, the compounds disclosed herein may be used to treat cancer or to inhibit the development of
10 cancer. The compounds are preferably administered either prior to or following primary treatment of tumours such as surgical removal of the tumours, treatment by administration of radiotherapy and/or conventional chemotherapeutic drugs or any other mode of treatment of the respective cancer or its precursors.

In passive immunotherapy, treatment involves the delivery of biologic reagents
15 with established tumour-immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumour effects and does not necessarily depend on an intact host immune system. Examples of effector cells include T lymphocytes (for example, CD8+ cytotoxic T-lymphocytes, CD4+ T-helper, tumour-infiltrating lymphocytes), killer cells (such as Natural Killer cells,
20 lymphokine-activated killer cells), B cells, or antigen presenting cells (such as dendritic cells and macrophages) expressing the disclosed antigens. The polypeptides disclosed herein may also be used to generate antibodies or anti-idiotypic antibodies (as in U.S. Pat. No. 4,918,164), for passive immunotherapy.

The predominant method of procuring adequate numbers of T-cells for adoptive
25 transfer immunotherapy is to grow immune T-cells in vitro. Culture conditions for expanding single antigen-specific T-cells to several billion in number with retention of antigen recognition in vivo are well known in the art. These in vitro culture conditions typically utilize intermittent stimulation with antigen, often in the presence of cytokines, such as IL-2, and non-dividing feeder cells. As noted
30 above, the immunoreactive polypeptides described herein may be used to rapidly expand antigen-specific T cell cultures in order to generate sufficient number of cells for immunotherapy. In particular, antigen-presenting cells, such as dendritic-, macrophage- or B-cells, may be pulsed with immunoreactive polypeptides or

transfected with a nucleic acid sequence(s), using standard techniques well known in the art. For example, antigen presenting cells may be transfected with a nucleic acid sequence, wherein said sequence contains a promoter region appropriate for increasing expression, and can be expressed as part of a recombinant virus or
5 other expression system. For cultured T-cells to be effective in therapy, the cultured T-cells must be able to grow and distribute widely and to survive long term in vivo. Studies have demonstrated that cultured T-cells can be induced to grow in vivo and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (see, for example, Cheever, M., et
10 al, "Therapy With Cultured T Cells: Principles Revisited," Immunological Reviews, 157:177, 1997).

According to the present invention, sets of one or more tumor associated polypeptides may be employed to generate and/or isolate tumour-reactive T-cells, which can then be administered to the patient. In one technique, antigen-specific
15 T-cell lines may be generated by in vivo immunization with short peptides corresponding to immunogenic portions of the disclosed polypeptides. The resulting antigen specific CD8+ CTL or CD4+ T-helper cells clones may be isolated from the patient, expanded using standard tissue culture techniques, and returned to the patient.

20 Alternatively, peptides corresponding to immunogenic portions of the polypeptides of the invention may be employed to generate tumour reactive T-cell subsets by selective in vitro stimulation and expansion of autologous T-cells to provide antigen-specific T-cells which may be subsequently transferred to the patient as described, for example, by Chang et al. (Crit. Rev. Oncol. Hematol., 22(3), 213,
25 1996). Cells of the immune system, such as T-cells, may be isolated from the peripheral blood of a patient, using a commercially available cell separation system, such as CellPro Incorporated's (Bothell, Wash.) CEPRATE.TM. system (see U.S. Pat. No. 5,240,856; U.S. Pat. No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). The separated cells are stimulated with one or more
30 of the immunoreactive polypeptides contained within a delivery vehicle, such as a microsphere, to provide antigen-specific T-cells. The population of tumour antigen-specific T-cells is then expanded using standard techniques and the cells are administered back to the patient.

In another embodiment, T-cell receptors and/or antibodies specific for the polypeptides can be cloned, expanded, and transferred into other vectors or effector cells for use in adoptive immunotherapy.

In a further embodiment, syngeneic or autologous dendritic cells may be pulsed
5 with peptides corresponding to at least an immunogenic portion of a polypeptide disclosed herein. The resulting antigen-expressing and/or presenting dendritic cells may either be transferred into a patient, or employed to stimulate T-cells to provide antigen-specific T-cells which may, in turn, be administered to a patient. The use of peptide-pulsed dendritic cells to generate antigen-specific T-cells and
10 the subsequent use of such antigen-specific T-cells to eradicate tumours in a murine model has been demonstrated by Cheever et al, (Immunological Reviews, 157:177, 1997).

Monoclonal antibodies directed against tumor associated peptides presented on cellular membranes may according to the present invention also be used as
15 therapeutic compounds in order to diminish or eliminate tumours. The antibodies may be used on their own (for instance, to inhibit metastases) or coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides, differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include ⁹⁰Y, ¹²³I, ¹²⁵I, ¹³¹I, ¹⁸⁶Re, ¹⁸⁸Re, ²¹¹At, and ²¹²Bi.
20 Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers include phorbol esters and butyric acid. Preferred toxins include ricin, abrin, diphtheria toxin, cholera toxin, gelonin, Pseudomonas exotoxin, Shigella toxin, and pokeweed antiviral protein.

Pharmaceutical compositions useful in immuno-therapy according to the present
25 invention may comprise a set of one or more tumor associated polypeptides (or variants thereof), and a physiologically acceptable carrier. The pharmaceutical composition may, for example, comprise one or more of the inventive peptides derived from tumor associated proteins. In one embodiment of the present invention, the pharmaceutical composition may additionally comprise one or more
30 further peptides derived from other proteins, which show non-wild type expression in tumors. Such proteins may for example be cell cycle regulatory proteins, proteases, such as metallo-proteinases, signal transduction molecules, receptors,

etc. In one embodiment, the additional peptides are derived from cell cycle regulatory proteins such as p13,5, p14, p15, p16, p19, p21, p27, p53, Rb protein, osteopontine, grp, renal dipeptidase, id1, mcm5, mcm2, cdc6, MDM2 etc. The peptides disclosed herein may be used in combination of one or more further
5 peptides associated with cancer.

Examples for peptides, that are associated with cancer are disclosed in WO9904265A2, WO0149716A2, WO0055633A2 and WO0142792A2, which documents shall be incorporated herein by reference. One or more of the peptides disclosed in this documents may be used for therapeutic and or diagnostic
10 purposes in combination with one or more peptides disclosed herein.

The vaccines may additionally comprise a non-specific immune-response enhancer, wherein the non-specific immune response enhancer is capable of eliciting or enhancing an immune response to an exogenous antigen. Examples of non-specific-immune response enhancers include adjuvants, biodegradable
15 microspheres (e.g., polylactic galactide) and liposomes (into which the polypeptide is incorporated). Pharmaceutical compositions and vaccines may also contain other epitopes of tumour antigens, either incorporated into a fusion protein or present within a separate polypeptide.

In one embodiment of the invention, the pharmaceutical composition for use in
20 immunotherapy may comprise further immunogenic molecules, which are known to effectively elicit immune responses. By this way the immune response to the target molecules may be enhanced as described in: Benoit, J.; et al., "T cell defined tumor antigens", Current Opinion in Immunology 9, 1997, 684-693 and: Markwin P., et. al., "Identification of Peptides for Immunotherapy of Cancer. It is
25 still worth the effort", Critical Reviews in Immunology. 18, 1998, 7-27. Both cited articles shall be included here within by reference.

Alternatively, a pharmaceutical composition or vaccine suitable for immunotherapy according to the present invention may contain nucleic acids, that code for one or more tumor associated polypeptides according to the present invention. Nucleic
30 acids may for example include single-stranded (sense or antisense) or double-stranded molecules, and may be DNA (genomic, cDNA or synthetic) or RNA. RNA molecules comprise as well HnRNA (containing introns) as mRNA (not containing

introns). According to the present invention the polynucleotides may also be linked to any other molecules, such as support materials or detection marker molecules, and may, but need not, contain additional coding or non-coding sequences. The nucleic acid may be administered in a way that allows the polypeptides to be
5 generated in situ. Suitable expression systems are known to those skilled in the art. The expression of the polypeptides may for example be persistent or transient. In pharmaceutical compositions and/or vaccines, providing for in-situ expression of polypeptides, the nucleic acids may be present within any suitable delivery system known to those of ordinary skill in the art, including nucleic acid expression
10 systems, bacteria and viral expression systems. Appropriate nucleic acid expression systems comprise the necessary regulatory nucleic acid sequences for expression in the patient, such as suitable promoters, terminators etc.

Bacterial delivery systems may for example employ the administration of a bacterium that expresses an epitope of a cell antigen on its cell surface. In a
15 preferred embodiment, the nucleic acid may be introduced using a viral expression system such as e.g., vaccinia virus, retrovirus, or adenovirus, which may involve the use of a non-pathogenic, replication competent virus. Suitable systems are known to those of ordinary skill in the art and are disclosed, for example, in Fisher-Hoch et al., PNAS 86:317-321, 1989; Flexner et al., Ann. N.Y. Acad Sci. 569:86-
20 103, 1989; Flexner et al., Vaccine 8:17-21, 1990; U.S. Pat. Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Pat. No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; Berkner, Biotechniques 6:616-627, 1988; Rosenfeld et al., Science 252:431-434, 1991; Kolls et al., PNAS 91:215-219, 1994; Kass-Eisler et al., PNAS 90:11498-11502, 1993; Guzman et al., Circulation 88:2838-
25 2848, 1993; and Guzman et al., Cir. Res. 73:1202-1207, 1993.

In another embodiment, transgenic mammalian cells may be used for delivery and/or expression of the nucleic acids. The methods for producing nucleic acid constructs suitable for in-situ expression of polypeptides are known to those of skill in the art.

30 Furthermore, the nucleic acid may be administered as naked nucleic acids. In this case appropriate physical delivery systems, which enhance the uptake of nucleic acid may be employed, such as coating the nucleic acid onto biodegradable

beads, which are efficiently transported into the cells. Administration of naked nucleic acids may for example be useful for the purpose of transient expression within a host or host cell.

The pharmaceutical compositions used for immuno-therapy according to the present invention may be administered by any suitable way known to those of skill in the art. The administration may for example comprise injection, such as e.g., intracutaneous, intramuscular, intravenous or subcutaneous injection, intranasal administration for example by aspiration or oral administration. A suitable dosage to ensure the pharmaceutical benefit of the treatment should be chosen according to the parameters, such as age, sex, body weight etc. of the patient, known to those of skill in the art.

The type of carrier to be employed in the pharmaceutical compositions of this invention, will vary depending on the mode of administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a lipid, a wax and/or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and/or magnesium carbonate, may be employed. Biodegradable microspheres (e.g., polylactic glycolide) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Pat. Nos. 4,897,268 and 5,075,109. Any of a variety of immune-response enhancers may be employed in the vaccines of this invention. For example, an adjuvant may be included. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a nonspecific stimulator of immune response, such as lipid A, Bordetella pertussis or Mycobacterium tuberculosis. Such adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, Mich.) and Merck Adjuvant (Merck and Company, Inc., Rahway, N.J.).

The pharmaceutical compositions or immuno-therapeutic methods according to the present invention may be used for the treatment of degenerative disorders or cancers. For example the compositions and methods may be employed in the

therapy of diagnosed cancers in order to eliminate the tumour cells from the affected organism. As well primary tumours as metastases or disseminated tumour cells within an organism may be targets to the therapeutic compounds and methods disclosed herein.

- 5 Furthermore, the compositions and methods of the invention may be employed in the treatment of pre-neoplastic conditions. In this case, the pre-neoplastic cells or tissues may be directly addressed by the immuno-therapeutic compositions or methods, or may be hindered from evolving into neoplastic or dysplastic conditions. For example in this case, the pre-neoplastic condition may be treated
10 preventively. By vaccination, the immune response may be elicited so that emerging neoplasms may be destroyed.

The methods and compositions according to the present invention may also be used for the prevention and therapy of tumors. For this purpose, a vaccination of a population of organisms or of subgroups of said population may be performed.

- 15 The subgroups may be built by suitable parameters such as hereditary predisposition for the emergence of degenerative disorders, exposure to factors that increase the risk of being affected by said disorders, etc.

- The present invention provides compositions and methods for enhanced detection and treatment of tumors. The invention provides peptides derived from proteins,
20 showing an altered expression in association with tumors. These peptides may thus be employed as a preventive vaccine, immuno-therapeutic agents against said disorders or may serve as diagnostic tools. Furthermore, the invention also provides sets of (poly)peptides useful for the treatment of particular types of disorders. The use of sets of tumor associated polypeptides for the immuno-
25 therapy according to the present invention provides the means for a reliable treatment of tumors reducing the risk of escape of several tumour cells or of a population of tumour cells from being addressed by the therapy. Thus, the method of the invention reduces the risk of recurrence of tumours after immuno-therapy employing CTLs, T-helper cells and possibly specific antibody producing B-cells
30 raised against tumor associated polypeptides characteristic for tumour cells.

The INK4a gene locus as used in the context of the present invention is located on chromosome 9p21. The locus is alternatively nominated e.g. the INK4a/ARF locus,

CDKN2A gene, the MTS1 locus. The gene locus dealt with herein is that gene locus, that in one aspect encodes the gene product p16INK4a protein, but is also known to encode for other gene product in alternative reading frames.

A gene product useful according to the present invention may be any molecule
5 transcribed from a gene or any molecule translated from such a transcript. Thus gene product as used in the context of the present invention may comprise polynucleotides such as e.g. DNA or RNA and polypeptides such as proteins, proteoglycans, peptides etc. Expression product as used in the context of the present invention shall comprise any transcript of a gene locus in forward or reverse
10 direction including any reading frames, splicing variants. Expression products as used herein shall thus comprise any alternative products encoded by the nucleic acids of a particular gene locus.

In one embodiment of the invention, the cell cycle regulatory proteins are encoded by the INK4a gene. The cell cycle regulatory proteins as disclosed herein comprise
15 any proteins encoded by alternative reading frames, arising from alternative splicing or usage of alternative promoter sequences of said gene. In one embodiment of the present invention the proteins exhibit molecular weights of about 5 to 40 kDa or any value in between and preferably of about 10 to 20 kDa or any value in between and most preferably of about 14 to about 19 kDa or any value in between respectively.

20 Discrimination as used in the context of the present invention shall comprise an assessment whether a sample is to be classified in one or another way. In a preferred embodiment of the invention the discrimination pertains to the assessment of a tissue or components thereof being neoplastic or being metaplastic. Thus the discrimination as used herein is a judgement about the growth properties of cells in
25 a sample.

Neoplastic as used in the context of the present invention shall refer to dysplasias from mild to severe dysplasias and their precursory stages, as well as carcinoma such as carcinomas in situ or invasive carcinomas and disseminated tumor cells. Neoplastic as used herein shall comprise early and precursor stages of dysplasias
30 and carcinomas. Metaplasia according to the method disclosed herein shall refer to

a state during the differentiation of cells being intermediate between two differentiated states.

The discrimination according to the present invention is based on the presence or absence of cells expressing p14ARF and on the presence or absence of cells
5 overexpressing p16INK4a in said sample. The cells expressing p14ARF need not to be the same cells as those overexpressing p16INK4a although the expression of both marker molecules may occur in the same cells.

Thus, the presence of cells expressing p14ARF gene-products in a sample simultaneously with the presence of cells overexpressing p16INK4a (other cells or
10 the same cells coexpressing both markers) according to the present invention serves to discriminate neoplastic or preneoplastic lesions from metaplasias.

INK4a encoded gene-products as used in the context of the present invention shall be any mRNA transcribed from the INK4a gene locus or any polypeptide translated from such an mRNA. INK4a gene-products suitable for the method according to the
15 present invention are gene-products such as e.g. p16INK4a and p14ARF.

The method for detection of the level of the INK4a encoded gene-products according to the present invention is any method, which may (but need not) be e.g. suited to detect even very small amounts of specific biological molecules in biological samples. The detection reaction according to the present invention is a
20 detection either on the level of nucleic acids or on the level of polypeptides.

The INK4a gene-products may be detected using reagents that specifically recognise these molecules. The detection reaction for the INK4a gene-products may comprise one or more reactions with detecting agents either recognizing the initial marker molecules or recognizing the prior molecules used to recognize other
25 molecules.

The detection of the different gene products may be performed in one reaction vessel or containment or in different containments simultaneously or subsequently in time. Thus the different gene products may be detected simultaneously in one cell coexpressing both products. Otherwise cell coexpressing the gene products may be

used for separated detection reaction (separated in space or in time) to detect each a single marker in the cells. In another embodiment there might be cells expressing one or the other marker. The detection of the marker molecules in the different cells may as well be performed simultaneously or separately in time and/or space.

- 5 The detection reaction further may comprise a reporter reaction indicating the presence or absence and/or the level of the INK4a gene-products. The reporter reaction may be for example a reaction producing a coloured compound, a bioluminescence reaction, a fluorescence reaction, generally a radiation emitting reaction etc.
- 10 In a preferred embodiment, different marker molecules may be recognized by agents that produce different reporter signals, so that the signals referring to marker molecules could be distinguished. In one preferred embodiment of the invention the detection of the expression of the two or more INK4a gene-products is carried out simultaneously. In this case the reporter reaction may for example employ different
15 fluorescent labels for the different molecules detected.

Applicable formats for the detection reaction according to the present invention may be, blotting techniques, such as Western-Blot, Southern-blot, Northern-blot. The blotting techniques are known to those of ordinary skill in the art and may be performed for example as electro-blots, semidry-blots, vacuum-blots or dot-blots.

- 20 Amplification reaction may also be applicable for the detection of e.g. nucleic acid molecules.

- In one preferred embodiment of the invention, the detection of the level of INK4a gene-products is carried out by detection of the respective mRNA or fragments thereof present in the sample. The means for detection of nucleic acid molecules
25 are known to those skilled in the art. The procedure for the detection of nucleic acids can for example be carried out by a binding reaction of the molecule to be detected to complementary nucleic acid probes, proteins with binding specificity for the nucleic acids or any other entities specifically recognizing and binding to said nucleic acids.

Probes as used in the context of the present invention may be any agent binding specifically to a molecule. In the case of nucleic acids a probe may be a oligonucleotide hybridising to a particular sequence. In one embodiment the probe may be e.g. a primer.

- 5 This method can be performed as well in vitro as directly in situ for example in the course of a detecting staining reaction. Another way of detecting the INK4a mRNAs in a sample performed in the method according to the present invention is an amplification reaction of nucleic acids, which can be carried out in a quantitative manner such as for example the polymerase chain reaction. In a preferred
10 embodiment of the present invention real time RT PCR may be used to quantify the level of INK4a mRNAs in samples of tumors.

In another preferred embodiment of the invention, the detection of the level of INK4a gene-products is carried out by determining the level of protein expression. The determination of the INK4a gene-product on the protein level can for example be
15 carried out in a reaction comprising a binding agent specific for the detection of the particular INK4a polypeptide.

The binding agents can be used in many different detection techniques for example in western-blot, ELISA or immuno-precipitation. Generally polypeptide binding agent based detection can be carried out as well in vitro as directly in situ for example in
20 the course of an immuno-cytochemical staining reaction. Any other method for determining the amount of particular polypeptides in biological samples can be used according to the present invention.

Binding agents as used in the context of the present invention for the detection of the level of INK4a polypeptides such as p16INK4a or p14ARF polypeptides may
25 comprise antibodies and antigen-binding fragments, bifunctional hybrid antibodies, peptidomimetics containing minimal antigen-binding epitopes etc.

An antibody or antigen-binding agent is said to react specifically, if it reacts at a detectable level with a protein disclosed herein, and does not significantly react with other proteins. The antibodies according to the present invention may be
30 monoclonal or polyclonal antibodies. As used herein, the term antibody or

monoclonal antibody is meant to include intact molecules as well as antibody fragments. Moreover, antibodies of the present invention include chimeric, single chain, and humanized antibodies.

According to the present invention, binding agents may be used isolated or in
5 combination. By means of combination it is possible to achieve a higher degree of sensitivity. The term antibody, preferably, relates to antibodies that consist essentially of pooled monoclonal antibodies with different epitopic specificities, as well as distinct monoclonal antibody preparations.

Monoclonal antibodies are made from antigen containing fragments of the
10 polypeptide of the invention using any of a variety of techniques known to those of ordinary skill in the art; see, e.g., Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, 1988. In one such technique, an immunogen comprising the antigenic polypeptide or a synthetic part thereof is initially injected into any of a wide variety of mammals (e.g., mice, rats, rabbits,
15 sheep and goats). In this step, the polypeptides of this invention may serve as the immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according
20 to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

The INK4a gene-products may according to the present invention be detected
25 simultaneously. In this context, simultaneously according to the present invention shall mean either literally at the same instant or within the same testing procedure, whereby the single detection steps are temporarily consecutive.

A sample according to the method of the present invention may comprise any sample comprising cells of anogenital origin. Samples may comprise e.g.
30 secretions, smears, body fluids, and cell-samples.

In one embodiment of the present invention, samples comprise cells of the uterine cervix. In a preferred embodiment of the present invention, the sample of cervical cells may be prepared according to a classical Pap smear. In a further preferred embodiment of the present invention the sample may be prepared as a monolayer
 5 or thin layer preparation of the cytological specimen.

Preparation of a sample may comprise e.g. obtaining a sample of a tissue, of a body fluid, of cells from a patient. According to the present invention, preparation of the sample may also comprise several steps of further preparations of the sample, such as preparation of dissections, spreading or applying the cells to be examined onto
 10 microscopic slides, preparation of tissue arrays, isolation of polypeptides or nucleic acids, preparation of solid phase fixed peptides or nucleic acids or preparation of beads, membranes or slides to which the molecules to be determined are coupled covalently or non-covalently.

The neoplastic lesions to which the method according to the present invention may
 15 be applied comprise any anogenital lesion, which is characterized by the overexpression of INK4a gene products. In one preferred embodiment of the present invention the anogenital lesion is a lesion of the uterine cervix.

Another aspect of the present invention is a testing kit for performing the method according to the present invention. The kit may be for example a diagnostic kit or a
 20 research kit.

A kit according to the present invention comprises at least an agent suitable for detecting the INK4a gene-products.

Thus a kit according to present invention may comprise:

- a) reagents for the detection of the INK4a gene-products
- 25 b) reagents and buffers commonly used for carrying out the detection reaction, such as buffers, detection-markers, carrier substances and others
- c) INK4a gene product samples for carrying out a positive control reaction

The reagents for the detection of the INK4a gene-products may include any agent capable of binding to the INK4a gene-products. Such reagents may include proteins, polypeptides, nucleic acids, peptide nucleic acids, glycoproteins, proteoglycans, polysaccharides or lipids.

- 5 The INK4a gene-product samples for carrying out a positive control may comprise for example nucleic acids in applicable form, such as solution or salt, peptides in applicable form, tissue section samples or positive cells.

In a preferred embodiment of the invention, the detection of the INK4a gene-products is carried out on the level of polypeptides. In this embodiment, the binding
10 agents may be for example antibodies specific for the INK4a gene-products or fragments thereof.

In another embodiment of the test kit, the detection of the INK4a gene-products is carried out on the nucleic acid level. In this embodiment of the invention, the reagent for the detection may be for example a nucleic acid probe or a primer reverse-
15 complementary to said INK4a nucleic acids.

The present invention provides a method for the discrimination of neoplastic and/or dysplastic and preneoplastic anogenital lesions, identifiable by assessment of the overexpression of p16INK4a, from metaplastic cells, which may also detectably express p16INK4a, in the course of cytological testing procedures. The method is
20 based on the detection of expressed gene-products of two or more INK4a gene products.

Thus the problem to be solved was to provide a method for discrimination between neoplastic and metaplastic cells especially in early stages of neoplasias, when cytological diagnostic methods based on the p16INK4a overexpression needs a
25 further information for the identification of metaplastic cells.

Furthermore the present invention provides a kit for performing the method according to the present invention.

EXAMPLES

The following examples are given for the purpose of illustration only and are not intended to limit the scope of the invention disclosed herein. For the purpose of illustration, the methods disclosed herein are exemplified using histological
 5 preparations. The histological examples aid to judge whether the cells stained in the one or the other way are to be classified as neoplastic or metaplastic. The methods may easily be transferred to cytological specimens by altering the protocol in appropriate manner. These alterations are known to those of ordinary skill in the art.

10 Example 1: **Immunochemical detection of the overexpression of p16INK4a and p14ARF in samples of the uterine cervix**

Sections of formalin fixed, paraffin embedded tissue samples of the cervix uteri were immunocytochemically stained using antibodies specific for p16INK4a and p14ARF.

The sections were rehydrated through incubation in xylene and graded ethanol, and transferred to Aqua bidest. Antigen retrieval was carried out with 10mM citrate buffer
 15 (pH 6.0) for p16INK4a. Thereafter, the slides were heated in a waterbath for 40 min at 95 °C. For p14ARF the slides were heated in 1mmol EDTA (pH8.0) at 98°C for 20min. The slides were cooled down to room temperature (RT) for 20 minutes, transferred to washing buffer (PBS / 0.1% Tween20) and finally the tissue sections were surrounded with a lipid-pencil.

20 For inactivation of endogenous peroxidase, the samples were incubated with 3% H₂O₂ for 20 min at RT and afterwards washed in PBS / 0.1% Tween20 for 5 to 10 min.

The slides were then incubated with the primary antibody, mouse anti-human p16INK4a antibody (1:25) for 30 min at RT. The slides were then rinsed with
 25 washing buffer and placed in a fresh buffer bath for 10 min. Excess buffer was tapped off and the specimen was covered with 100µl of visualization reagent for 30 min at RT. Slides were washed as before, and covered with 200 µl substrate-chromogen solution (DAB) for 10 min. Then slides were washed as before and counterstained for 3 min in a bath of hematoxylin. Residual hematoxylin was rinsed

with distilled water, and specimens were mounted and coverslipped with an aqueous mounting medium.

Staining with the mouse anti-human p14ARF antibody was done as described before, except the antibody dilution was 1:100 and the incubation time was 1 h at

5 RT.

The microscopic examination of the slides reveals that cells immunoreactive with p16INK4a together with cells immunoreactive with p14ARF protein were only found in samples that might microscopically be identified as samples of neoplastic lesions. Cells stained by the p16INK4a specific reaction, which were originating from
10 metaplasias, were not stained by the reaction specific for the p14ARF protein. The microscopic inspection of the p14ARF stained slides shows that metaplastic cells were not immunoreactive with the antibodies directed against p14ARF protein. Samples containing dysplastic tissue areas in contrast comprised cells, which were immunoreactive with antibodies directed against p14ARF and antibodies directed
15 against p16INK4a. In contrast to dysplasias in metaplasias, no cells were stained using the p14ARF specific antibody.

The results show that the staining with reagents specific for p14ARF allows to discriminate p16INK4a overexpressing metaplasias from dysplasias.

20 **Example 2: Detection of cells expressing p14ARF or p16INK4a in samples of the uterine cervix by in situ hybridization**

Smears of the uterine cervix can be semi-quantitatively analysed for the mRNA level of p16INK4a and p14ARF in an in-situ staining reaction. The staining reaction is performed as follows:

For rehydration the spray-fixed smears are incubated in fresh 50% EtOH on a
25 rocking device. The PEG film produced by the fixation procedure is removed by intensive rinsing. Then the smears are rinsed in aqua bidest. The smears are incubated with porteinase K (10µg/ml in PBS) for 10 min at 37°C. Then the slides are transferred to washing buffer (PBS / 0.1% Tween20) and finally the area containing the cells is surrounded with a lipid-pencil.

The hybridization mixture is prepared by mixing 50 µl of ready to use hybridization buffer (DAKO A/S, Glostrup, Denmark) with about 5 – 10 pmol of the probes. The probes are fluorescein-labelled oligonucleotides of sequences complementary to the respective mRNAs.

- 5 The hybridization mixture is heated to 95°C and afterwards equilibrated to 37°C. After the boiling procedure, the smears are incubated with each 50 µl of the hybridization mixture for 4 hours at 42°C. The samples are washed in excess volumes of the wash buffers two times in 2 x SSC at 37°C for 15 min and once in 1 x SSC at 37 °C for 15 min. Then the smears are rinsed two times at room
- 10 temperature in 2 x SSC. Following this washing procedure, the dissections are incubated for 30 min with a blocking buffer (NEN, Blockingbuffer) at room temperature. Then follows 1 hour incubation with a 1:100 diluted (in Blocking buffer, see above) anti-Fluorescein-alkaline phosphatase (DAKO A/S). The smears are then washed 2 times in 1 x PBS/0,1% Triton X-100 for 10 min at room temperature,
- 15 followed by one wash step with 1 x PBS, 50 mM MgCl₂ (pH 9.2) for 10 min at room temperature. Then the staining reaction is performed with NBT/BCIP (Sigma) for 30 min to 2 hours at room temperature. The staining reaction is stopped by a short incubation with 1 mM EDTA in PBS. Finally the smears are dipped in distilled H₂O. and embedded with AquaTex (Merck). Then the stained dissections can be
- 20 analyzed microscopically.

Microscopic analysis reveals that metaplasias comprise cells expressing p16INK4a but no cells expressing p14ARF mRNA at detectable levels. In dysplastic lesions of the cervix uteri, cells expressing p16INK4a can be found and furthermore cells expressing p14ARF mRNAs at detectable levels can also be found.

- 25 This result indicates that the method according to the present invention may be used for the discrimination of metaplasias and neoplastic lesions.

Example 3: **In vitro stimulation of cellular immune response by HLA-A1 restricted peptides**

The present experiments are performed in order to determine whether the
5 peptides according to the present invention are suited to stimulate a cellular immune response. The experiments are performed as follows:

Peptides displaying HLA-A2.1-as well as HLA-A1 and HLA-A3-binding motifs are selected by taking advantage of specific computer programs [(Parker, Bednarek, & Coligan 1994); http://bimas.dcrt.nih.gov/molbio/hla_bind/ and (Rammensee et al.
10 1999); <http://134.2.96.221/scripts/MHCServer.dll/home.htm>]. Peptides are purchased from the peptide synthesis unit of the DKFZ. Stock solutions (10mg/ml in DMSO) are stored at -70°C and diluted to 1mg/ml in PBS before use.

In one representative experiment, T2 cells are pulsed with 7 p14ARF specific HLA-A2.1 restricted nonamer peptides (Pos. 51: MVRRLVTL, Pos. 91: AVALVLMML,
15 Pos. 92: AVALVLMML, Pos. 97: LMLLSQRL, Pos. 8: HIMGRGRCV, Pos. 85: AAPGAPAAV, Pos. 87: GAPAAVALV; 50µg/ml peptide) and 5µg/ml β2-microglobulin (Sigma; Deisenhofen, Germany) overnight at 37°C. The expression of HLA-A2.1 is then analyzed by flow cytometry using mAb BB7.2, followed by incubation with FITC-conjugated F(ab')₂ goat anti-mouse Ig (Vonderheide et al.
20 1999). Peripheral blood is obtained from healthy HLA-A2.1 donors and collected in heparinized tubes. PBMC are isolated by Ficoll-density gradient centrifugation. Whole CD3⁺ T cells are isolated from PBMC by magnetic depletion of non-T cells using the MACS Pan T Cell Isolation Kit (Miltenyi; Bergisch Gladbach, Germany) according to manufacturer's instructions. Preparations contain at least
25 97% of CD3⁺ cells as assessed by immunophenotypic analysis.

CD40 Bs of a HLA-A2.1 donors are incubated with the same peptides listed above (10µg/ml) and human β2-microglobulin (3µg/ml; Sigma) in serum-free Iscov's DMEM medium for one hour at room temperature, washed twice to remove excess of peptide, irradiated (30Gy) and added to purified CD3⁺ autologous T cells (>97%
30 CD3⁺) at a ratio of 4:1 (T:CD40 Bs) in Iscov's MEM containing 10% human AB-serum, supplements (1:100), and hIL-7 (10ng/ml, R&D). Cells are plated at a density of 2 x 10⁶ T cells/well in 1ml of medium. After three days in culture they are

fed with 1ml complete medium. For restimulation of T cells, this is repeated weekly. IL-2 is first given at day 21 (10IU/ml, R&D), and also at day 24. From day 28 on, only hIL-2 is used instead of hIL7.

ELISpot assays are performed as described elsewhere (Meyer et al. 1998). Briefly, 5 nitrocellulose-96-well plates (Multiscreen; Millipore, Bedford, USA) are covered with mouse anti-human IFN-g monoclonal antibody (Mabtech, Sweden) and blocked with serum containing medium. Varying numbers of effector cells are plated in triplicates with $3,5 \times 10^4$ peptide-loaded T2 cells per well as targets. After incubation for 18h, plates are washed, incubated with biotinylated rabbit anti- 10 human IFN-g second antibody, washed again, incubated with streptavidin coupled alkaline phosphatase, followed by a final wash. Spots are detected by incubation with NBT/BCIP (Sigma) for 45min. The reaction is stopped with water. After drying, spots are counted using the KS-ELISpot reader (Zeiss Kontron; Göttingen, Germany).

15 The analysis shows that the used peptides are suited to raise an immune response. Immunogenic peptides deduced from the p14ARF sequence thus may be used to raise immune response for example in the course of vaccinations according to the present invention.

20 Example 4: **Cytotoxicity assay directed against cells displaying peptides.**

CTL bulk cultures and/or CTL clones obtained according to the method described in Example 1 are tested for their cytotoxicity as follows:

To identify the cytolytic potential of CTL clones, one HLA matched HLA-A2.1 positive cancer cell line (CASKI) expressing p14ARF as well as p14ARF 25 transfected, p14ARF negative cell line (SW480) of the same HLA haplotype are analysed.

After selection and expansion of the transfected cell lines, at least two stably transfected sub-cell lines are available. These cell lines are used in cytotoxicity assays, wherein negative controls are p14ARF negative cell lines (SW480, 30 HCT116), one p14ARF positive cell line of different HLA-haplotype (SiHA) and/or the respective untransfected cell line (SW480). It can be shown that the

transfected cell lines as well as p14ARF positive cell lines of the correct haplotype are lysed by the bulk cultures and/or pooled clones of CTLs. The reactivity is tested at different target cell to effector cell ratios. In average, around 20%-30% of the target cells are lysed in the assays. The control cells are always lysed at a
 5 significantly lower percentage.

These results are representative for the results related to the other peptides (HLA-A1 and HLA-A3 restricted), which render similar rates of lysis.

The experiments show that the peptides disclosed herein may raise immune response. The peptides may furthermore be applied for the detection of the
 10 presence of cytotoxic T-cells directed against a particular frameshift peptide.

Example 5: Screening for Antibodies directed against peptides derived from INK4a encoded proteins in patient samples

Sera of 25 patients with diagnosed cervical dysplasias are tested for the presence
 15 of antibodies to peptides derived from p14ARF as disclosed herein. As a control, sera of 50 normal individuals are tested.

For the test, synthetic peptides representing immunogenic regions of the p14ARF protein sequence associated with cervical dysplasias are spotted onto nylon membranes. The nylon membranes are subsequently incubated for one hour in
 20 phosphate-buffered saline (PBS) with 5% milk powder for blocking unspecific membrane binding. After washing the membranes 3x with PBS, the membranes are incubated with the test and control sera. The sera are diluted 1:1,000 in PBS/0,5% milk powder and incubated overnight with gentle shaking. Subsequently the sera are removed, and membranes are washed three times in PBS before they
 25 are incubated with a polyclonal alkaline phosphatase conjugated goat anti-human IgG antibody for one hour. Thereafter, the membranes are washed repeatedly with PBS/0,05% TWEEN-20 before staining reaction is developed using nitroblue tetrazolium chloride and bromochloro-indoyl-phosphate (SigmaAldrich) in Tris-buffered saline (TBS). Binding of human antibodies specific for individual p14ARF
 30 specific polypeptides thus is made visible by color-deposit on the respective membrane.

The results show that in samples of tumor patients, antibodies directed against at least one p14ARF specific peptide are present.

This illustrates that the method according to the present invention may be used for diagnosis of tumors such as cervical dysplasias.

5

Example 6: Immunoenzymatic detection of the over-expression of p16INK4a and p14 ARF in cytological samples of the uterine cervix

Merckofix® fixed cytological samples (conventional smears and liquid based cytology (ThinPreps®)) of the cervix uteri were immunoenzymatic double stained
10 using antibodies specific for p16INK4a and p14ARF.

Conventional smears and liquid based cytological samples were rehydrated in ethanol (50%) for 10min at RT and transferred in Aqua bidest. Antigen retrieval was carried out with 1mM EDTA buffer (pH 8.0) for p16INK4a and p14ARF.

Thereafter the slides were heated in a waterbath for 40 min at 95 °C – 98°C. The
15 slides were cooled down to RT for 20 minutes and transferred to washing buffer.

Endogenous peroxidase activities were blocked with 3% H₂O₂ for 5min at RT.

After washing, the slides were incubated with the first primary antibody, mouse anti-human p14ARF antibody, overnight at 4°C and were then rinsed with a washing buffer and placed in a fresh buffer bath for 5 min. Excess buffer was
20 tapped off and each specimen was covered with 200µl of the secondary reagent (EnVision goat anti mouse – Peroxidase) and incubated for 30min at RT . Then slides were washed three times as before. For the chromogenic visualization, DAB (DakoCytomation) was used by incubating the slides with the substrate chromogen complex for 10 min at RT. The reaction was stopped in deionized
25 water and slides placed in wash buffer.

After washing, the slides were incubated with the second primary antibody directed against p16^{INK4a} (FITC labeled) for 30 min at RT and were then rinsed with washing buffer and placed in a fresh buffer bath for 5 min. Excess buffer was tapped off and each specimen was covered with 200µl of the secondary reagent

(rabbit anti FITC – Alkaline Phosphatase labeled / DakoCytomation) and incubated for 30min at RT . Then slides were washed three times as before. For the chromogenic visualization, Fast Red (BioGenex) was used by incubating the slides with the substrate chromogen complex for 30min at RT. The reaction was
 5 stopped in deionized water .

After counterstaining with hematoxylin (DakoCytomation) for 2min at RT, the slides were incubated in running tap water for 10min at RT and then mounted with an aqueous mounting medium (Aquatex / MERCK).

The microscopic examination of the slides revealed that cells immuno-reactive
 10 with p16INK4a and also immuno-reactive for p14ARF could be identified as dysplastic cells on the basis of their morphology. Cells stained by the p16INK4a specific reaction, which were originating from metaplasias, were not stained by the reaction specific for p14ARF. Dysplastic cells in contrast were immuno-reactive with p14ARF and with antibodies directed against p16INK4a. (cf. Figure 5). So in
 15 contrast to metaplastic cells, in dysplastic cells, double-staining of single cells could be produced using the p14ARF and p16INK4a specific antibodies.

The results as given in Figure 5 show that double staining of cells with reagents specific for p14ARF and p16INK4a allows for double staining of dysplasias in immunoenzymatic double staining procedures.